

For Professional Use Only For Emergency Use Authorization (FUA) Only For In Vitro Diggnostic Use

REF Test strips and swabs L016000609024,L016000609048 **REF** Test strips no swabs L016000109012, L016000109024, L016000109048

IVD Rx Only

Date of Povision 2023 08

SDEC 20211 D10 ADTO0570 D12 LumiraDx SARS-CoV-2 Ag Test

(Aq) Test Strips (hereafter referred to as Test Strips) are to be used with the LumiraDy Platform The LumiraDy Platform is a point of care system for professional use which is used for in vitro diagnostic tests. It comprises a This test is for HEALTHCADE PROFESSIONAL USE ONLY and allows users to perform tests using small sample volumes and to view results quickly on the Instrument touchscreen

The LumiraDx SARS-CoV-2 Ag Test is a rapid microfluidic immunofluorescence assay for use with the LumiraDx Platform intended for the auglitative detection of the nucleocapsid protein antigens from SAPS. V-2 in direct anterior nasal swab and nasopharvnaeal swab specimens collected by a healthcare provider from individuals who are suspected of COVID-19 within the first twelve (12) days of symptom onset when tested at least twice over three days with at least 48 hours between tests and from COVID-19 when tested at least three times over five days with at least 48 hours between tests

Testing is limited to laboratories certified under the Clinical Laboratory mprovement Amendments of 1988 (CLIA), 42 U.S.C 263a, that meet the requirements to perform moderate high or waived complexity tests This product is authorized for use at the Point of Care (POC), i.e. in patient care settings operating under a CLIA Certificate of Waiver, Certificate of Compliance, or Certificate of Accreditation

The LumiraDx SARS-CoV-2 Aa Test does not differentiate between SARS-CoV and SARS-CoV-2

Results are for the identification of SARS-CoV-2 nucleocapsid protein antigen, which is generally detectable in anterior nasal swab and asopharynaeal swab specimens during the acute phase of infection Positive results indicate the presence of viral antigens but clinical necessary to determine infection status. Positive results do not rule out bacterial infection or co-infection with other viruses. The agent detected may not be the definitive cause of disease. Laboratories within the United States and its territories are required to report all results to the appropriate public health authorities All negative results are presumptive and confirmation with a molecular

assay, if necessary, for patient management, may be performed. Negative results do not rule out SARS-CoV-2 infection and should not be used as the sale basis for treatment or nationt management decisions including infection control measures such as isolating from others and wearing masks. Negative results should be considered in the context of a patient's recent exposures, history and presence of clinical signs and symptoms consistent with COVID-19

The LumiraDx SARS-CoV-2 Ag Test is intended for use by medical professionals or operators who are proficient in performing tests in point of

The LumiraDy SARS-CoV-2 Ad Test is only for in vitro diagnostic use under the Food and Drug Administration's Emergency Use Authorization This product has not been FDA cleared or approved.

Caution: For in vitro diagnostic use



Before you start testing, if you are new to the LumiraDx Instrument and LumiraDx Platform, you must read the LumiraDx Platform User Manual, the LumiraDx SARS-CoV-2 Aa Test Quick Reference Instructions and this entire Product Insert. In addition please watch the LumiraDx Platform Training Video available at lumiradx.com

Summary and explanation of the Test

by SARS-CoV-2 virus as coronavirus 2019 or COVID-191 The most common symptoms of COVID-19 are fever tiredness and dry cough. Some patients may have aches and pains, nasal congestion, headache, conjunctivitis, sore throat, diarrhea, loss of taste or smell, or a rash on skin or discoloration of fingers or toes. These symptoms are usually mild and begin gradually. Some people become infected but do not develop any symptoms and do not feel unwell. However, the disease can develop rapidly and have high morbidity in certain populations, especially those with underlying healt conditions. The disease can spread from person to person through small droplets from the nose or mouth which are spread when a person with COVID-19 coughs or exhales. Most estimates of the incubation period for COVID-19 range from 2-14 days².

The use of a LumiraDx SARS-CoV-2 Aa Test will enable the physician to verify infection quickly, begin proper treatment and to initiate isolation precautions helping prevent further spread of infection

Principle of the assay ne LumiraDy SAR-CoV-2 Ad Test is a single use fluorescence immunoassa

device designed to detect the presence of the nucleocapsid protein

nasopharyngeal swab sample using a recommended swab which is

in Extraction Buffer is added to the Test Strip using the vial dropper cap

provided The LumiraDx Instrument is programmed to perform the tes

eluted into a vial containing Extraction Buffer A single drop of the sample

protocol using the dried reagents contained within the strip. The test result is

determined from the amount of fluorescence the Instrument detects within

the measurement zone of the Test Strip The concentration of the analyte

are displayed on the Instrument touchscreen within 12 minutes from the

LumiraDxTest Strips packed individually in sealed desiccant foil

PEID (Padio frequency ID) Tag held inside the Test Strip carton

LumiraDy SAPS-CoV-2 Ag Test Quick Peference Instructions

product codes L016000609024 L016000609048

Materials required but not provided with the Test Strip carton

and organisational compliance)

with the LumiraDx SARS-CoV-2 Aa test.

Sterile Nasal Collection Swabs (Provided only with the following

LumiraDx SARS-CoV-2 Ag Quality Controls (as required to meet local

Note: If using a LumiraDx SARS-CoV-2 Ag test kit which does not

include swabs please note that the swabs listed on lumirady com

Read all instructions carefully before performing the test. Failure

to follow the instructions may result in inaccurate test results.

In the USA, this product has not been FDA cleared or approved, but

inder the CIIA that meet the requirements to perform moderate

has been authorized by FDA under an Emergency Use Authorization

(EUA) for use by authorized laboratories; use by laboratories certified

high or waived complexity tests. This product is authorized for use at

the Point of Care (POC), i.e., in patient care settings operating under

This product has been authorized only for the detection of proteins

from SARS-CoV-2, not for any other viruses or pathogens.

The emergency use of this product is only authorized for the

duration of the declaration that circumstances exist justifying

the authorization of emergency use of in vitro diagnostic tests for

detection and/or diagnosis of COVID-19 under Section 564(b)(1) of

ne Federal Food, Drug, and Cosmetic Act, 21 U.S.C. § 360bbb-3(b)

(1), unless the declaration is terminated or authorization is revoked

Serial testing should be performed in individuals with pegative

at least 48 hours between tests) for asymptomatic individuals

If the patient has had symptoms longer than 12 days, consider

testing at least three times over five days with at least 48 hours

You may need to purchase additional tests to perform this serial

results at least twice over three days (with 48 hours between tests)

or symptomatic individuals and three times over five days (with

a CLIA Certificate of Waiver, Certificate of Compliance, or Certificate

may have only been internally validated by LumiraDx.

in the sample is proportional to the fluorescence detected. The results

antigen directly from SAPS-CoV-2 in either anterior pasal swah, or

The test procedure involves collecting an anterior pasal swab, or

nasonharvnaeal swah samples without transport media

addition of the sample

LumiraDy Test Product Insert

Extraction Buffer Vials

LumiraDy Instrument

Warnings and precautions

For in vitro diagnostic use

For prescription use only

(repeat) testing.

other materials.

Materials provided

- Exercise the normal precautions required for handling all laboratory reagents. Wear protective clothing such as laboratory coats, are collected and evaluated
 - when working with SARS-CoV-2 patient samples. Patient swabs, used Test Strips and used extraction buffer vials may be potentially infectious Proper handling and disposal methods should be ederal regulations.
- mall amounts however, should any reagent become exposed it should be treated as potentially infectious
- Be careful to minimize the risks of cross-contamination when testing nationt specimens, which can cause false positive results. Insufficient cleaning of the workspace, insufficient disinfection of the instrument, or inappropriate use of protective equipment (for example, failing to change aloves between patients) can increase the risk of cross contamination between specimens with subsequent false positive results. Consider the CDC guidance available at https://www. dc.gov/coronavirus/2019-ncov/lab/point-of-care-testing.html for changing gloves and cleaning work area between specimen handling and processing
- Avoid contact with your skin eyes nose or mouth. Do not ingest any kit components The extraction buffer may contain barmful. chemicals (see table below). If the solution contacts your skir

and organisational compilance)			
LumiraDx Connect if connectivity required (refer to LumiraDx Connect User Manual)	Chemical Name	Concentrations	GHS Code
Standard nasal or nasopharyngeal collection equipment is required	Hydrochloric acid	< 0.01%	H302, H315, H320
if using a LumiraDx SARS-CoV-2 Ag test kit which does not include swabs (L016000109012, L016000109024, L016000109048). Please visit	Sodium azide	0.09%	H302, H315, H320
lumiradx.com for information on LumiraDx validated swabs for use	- For more information	an an EUA a plagas visits ht	tas: / /v.s.s.vfda asv./

- emergencypreparedness-and-response/mcm-legal-regulatory-andpolicy-framework/emergencyuse-guthorization

Store the Test Strips in their original carton. You can store the Test Strips at a temperature between 2°C and 30°C (36°F and 86°F). Avoid freezing or storing in any area that could exceed 30°C. When stored properly the Test Strips can be used until the expiration date printed on the Test Strip foil pouch and the Test Strip carton. Discard the Test Strips if they are beyond

Handling the Test Strips

When you are ready to perform a test, open the Test Strip carton, take out a Test Strip and remove it from the fail pouch. Hold the Test Strip by gripping the blue label end with the label facing upward. Do not touch Test Strin mple Application Area. Do not bend or fold the Test Strip. Do not touc Strip contacts After removing the Test Strip from the foil pauch, it should be used immediately. Do not use the Test Strip if there are any visible signs of

- Anterior Nasal Swab Sample (NS)
- Nasopharvnaeal Swab Sample (NP)
- Rabbit and mouse monoclonal antibodies
- Magnetic particles Buffer and stabilizing agents
- Do not open the test strip until ready for immediate use. Discard and do not use any damaged or dropped Test Strips or
- Check the integrity of the individual swab packaging for damage. If damaged discard and do not use.
- Discard and do not use any damaged or dropped Nasal collection
- Do not use supplied Nasal swabs for Nasopharvnaeal sample To avoid sample contamination avoid touching the swab sampling
- head before and after sample collection. Inadequate or inappropriate sample collection, storage, and
- transport can result in incorrect results. The test cannot be visually interpreted; the LumiraDx Instrument must
- be used to generate results. Do not use the kit components beyond the expiration date
- Test components are single-use. Do not re-use. Do not mix components from different kit lots
- Samples must be processed as indicated in the Sample Extraction
- and Performing a Test sections of this Product Insert.
- All components of this kit should be discarded as Biohazard waste according to Federal, State and local regulatory requirements.

- Defer to the product refety data sheet for risk and refety phrases Lot Calibration File installation and disposal information. The product safety data sheet is available via our website at https://lumiradx.com/us-en/what-we-do/ diagnostics/test-technology/antigen-test
- disposable gloves safety mask and eve protection when samples Proper laboratory safety techniques should be followed at all times
- established by the laboratory in accordance with local state and Reagents encapsulated within the Test Strip are present in extremely
- eves nose or mouth flush with large amounts of water If irritation persists, seek medical advice; https://www.poisonhelp.org.or

	Chemical Name	Concentrations	GHS Code
d	Hydrochloric acid	< 0.01%	H302, H315, H320
t	Sodium azide	0.09%	H302, H315, H320
	For more information	on on ELIAs plagsa visit: ht	tos://www.fda.gov/

- For the most up to date information on COVID-19, please visit: www.

Storing the Test Strips

damage to the foil pouch such as tears or holes

The following samples can be used with the LumiraDx SARS-CoV-2 Ag Test

- Fluorescent particles

Preparing the Instrument to perform a Test

Power on the Instrument by pressing the power button at the rear of the Instrument. You will hear the Instrument powering on, and the display will be a blank black screen for several seconds before starting up. If the screen is just dimmed tap the touch-screen to wake up the Instrument.

Refer to the section on **Performing a Test** in this Product Insert for rmation on how to test a Patient sample. The LumiraDx Quick Reference Instructions (QPI) provide an illustrated step-by-step procedure on how to run a Test. Operate the LumiraDx Platform at room temperature between 15°C and 30°C (59°F and 86°F) and 10% - 75% relative humidity

The Instrument will prompt to install the Lot Calibration File when inserting a new Test Strip Lot Once installed the Instrument will have all the information required to process the test, and any future tests from the same Lot of Test

Lot Calibration Files are required to provide the Instrument with information needed to perform diagnostic tests. This only needs to be completed once for each Test Strip Lat The Instrument will prompt to install the Lot Calibration File when inserting a new Test Strip Lot.

displayed



When indicated by the touchscreen open the fail pouch just before use and insert the LumiraDx Test Strip into the LumiraDx Instrument The Instrument will indicate when it is ready for the sample to be applied. The LumiraDx SARS-CoV-2 Aa Test results should be evaluated by a Healthcare Professional in the context of all available clinical and laboratory

Instructions for sample collection

PFID strip code reader

Touch back of Test Strip

Carton $((\bullet))$ symbol to install.

and guidelines according to your organization. For collection of nasal swahs and nasonharyngeal swahs follow the Centers for Disease Control and Prevention (CDC) Swab Collection Guidelines and swab anufacturers' recommendations. Users should be trained in appropriate sample collection and handling procedures The steps that follow apply to an anterior nasal swab and nasopharynged

When collecting any type of sample, follow universal collection precautions

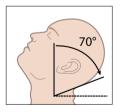
Swabs provided in the kit (L016000609024, L016000609048) For anterior nasal

information on LumiraDx validated swabs for use with the LumiraDx SARS-

sampling where swabs are provided please use the swabs within the kit. Swabs not provided in the kit (1016000109012 101600010902/ Where a swab is not provided within the kit, please visit lumiradx.com for

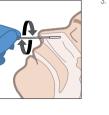
Sampling from an anterior nasal swab

CoV-2 Aa test







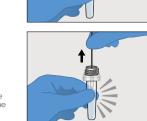


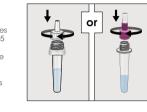
A swab sample is needed rom both nostrils, and this is taken using the same swab Remove swab from the swab packet. Hold the swab by the shaft, while gently rotating the swab, insert swab less than one inch into the first nostril until resistance is met at Turbinates (Turbinates are the small structures inside the nose).

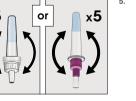
Tilt patient's head back 70°

Rotate the swab at least 4 times against the nasal wall for 10-15 seconds. Remove and repeat swab into the second nostril Then place the Swab into the Extraction Vial. See instruction for Sample Extraction

Sampling from a nasopharynaeal swab







Performing a Test (refer to the Quick Reference Instructions to make sure A positive test result means that the virus that equipes COVID 10 was that your Instrument has been prepared before starting this step). If using a frozen sample, the sample must be at room temperature before

Gently invert the Extraction Vial five times (5x) just before applying

Apply the extracted sample from the Extraction Vial onto the Sample Application Area of the inserted Test Strip To do this gently press the sides of the extraction vial until **one whole drop** is visible and allow it to touch the Sample Application Area of the Test Strip The sample will then be drawn by capillary action into the Test Strip are enabled) and a confirmation message will be displayed. The to uch creen of the Lumira Dy Instrument will request the user to immediately close the door (Note: you have 10 seconds only to Do not add more than one drop of sample. Do not open the

the sample to the Test Strip.

- door while the test is in progress The touchscreen will indicate test The result will appear on the Instrument touchscreen within 12
- minutes of applying the sample and starting the test. The results will be displayed as a positive or negative result SARS-CoV-2-Ag on the nstrument screen. (see Fig 1 and Fig 2). Dispose of the swab. Extraction Vial and Test Strip in the appropriate **Disinfection** of the Instrument with LumiraDx approved materials is
- nended if contamination is suspected and at least once per day when in use A list of approved disinfecting materials is available at lumirady com. Use the wine until the surface of the Instrument is visibly wet Allow the surface to remain wet for 1 minute and let air If you need to retest, you will use a new Test Strip. Use the same

extraction vial and repeat the test. The extracted sample must be

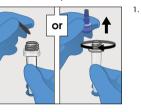
used within 5 hours of preparation when stored at room temperature

xtracted nasal and nasopharynaeal swab samples may be frozen

at -80°C and used up to 5 days after freezing After patient swabbing, process the Swab in the Extraction Vial as soon Interpretation of Results as possible. Do not place the swab back into the swab packaging sleeve Repeat testing is needed to improve test accuracy. Please follow the table

Instructions for sample extraction

after sample collection.



or blue screw cap from the top of the Extraction Vial containing the Extraction Buffer.

Place and soak the

Extraction Buffer for 10

seconds and then stir.

swab against the side

well by rotating the

of the vial 5 times

Firmly attach the

Dropper Lid to the top

must be used within 5

hours of preparation

room temperatur

Extracted nasal or

nasopharyngeal

be frozen at -80°C

and used up to 5

when stored at

of the Extraction Vial

clear or purple

Remove the seal

Tilt patients head back 70°

Pemove swah from the swah

nacket Hold the Swah by the

shaft firmly between the finger

to the palate until resistance is

encountered The Swah should

reach denth equal to distance

from postrils to outer opening

Gently rub and roll the Swah

ing it in place for several

a deviated sentum or blockage

creates difficulty in obtaining the

sample from one nostril use the

Slowly romayo the swab while

collected from both nostrils, but

it is not necessary if the swab

is saturated with fluid from the

first nostril. Remove and then

Extraction

place the swab in the extraction

vial. See instructions for Sample

rotating it Samples can be

from the other postril

same swab to obtain the sample 4

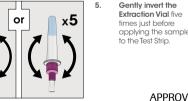
seconds to absorb secretions If

Swah through the nostril narallel

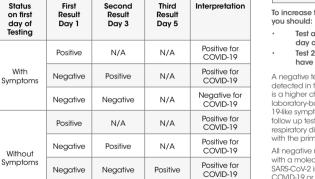
and gently and slowly insert



Sauceze Swah Swab while saueezina the middle of the Extraction Vial to remove the liquid from the swab. Discard the swab in biohazard waste.



below when interpreting test results.



Negative Negative Negative COVID-19

Results should be considered in the context of an individual's recent exposures, history, and the presence of clinical signs and symptoms consistent with COVID-19

→ © 100% (■■)

Patient Test Fini

COVID-19 Positive (+)

DOB: 07 Apr 1979 Gender: Male Patient ID: 123456789

POSITIVE +

SARS-CoV-2 Ag

Comment

result at any time.

OS-04277-01

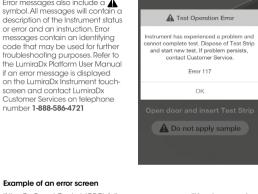
APPROVED FOR USE IN THE US.

EMERGENCY USE AUTHORIZED ONLY.

SARS-CoV-2 Ag

result screen display:

ouch-screen. Alert messages The result will be displayed on the Instrument screen - example of positive symbol. All messages will cor screen and contact LumiraDx number 1-888-586-4721



Patient Test

Repeat testing does not need to be performed if patients have a positive

Calibration file has not yet been loaded, at which point it will request it.

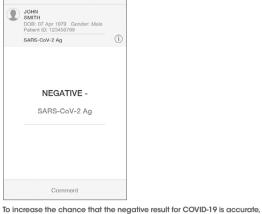
detected in the sample, and it is very likely the individual has COVID-19 and is contagious. Please contact the patient's doctor/primary care physician (if applicable) and the local health authority immediately and instruct your patient to adhere to the local quidelines regarding self-isolation. There is a very small chance that this test can give a positive result that is incorrect (a false positive) Positive results do not rule out bacterial infection or ac infection with other

viruses. The agent detected may not be the definite cause of disease. Individuals who test positive with the LumiraDx SARS-CoV-2 Aa test should self-isolate and seek follow up care with their physician or healthcare provider as additional confirmatory testing with a molecular test for positive results may also be necessary if there is a low likelihood of COVID-19 such as in individuals without known exposures to COVID-19 or residing in communities with low prevalence of infection COVID-19 Negative (-)

The result will be displayed on the Instrument screen - example of

Patient Test Fir

negative result screen display: □ 100% (
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Test again in 48 hours if the individual has symptoms on the first

- day of testing. Test 2 more times at least 48 hours apart if the individual does not have symptoms on the first day of testing.
- A negative test result indicates that the virus that causes COVID-19 was not detected in the sample. A negative result does not rule out COVID-19. There a higher chance of false negative results with antigen tests compared to oratory-based tests such as PCP tests. If the test is negative but COVID-19-like symptoms, e.a., fever, cough, and/or shortness of breath continue follow up testing for SARS-CoV-2 with a molecular test or testing for other ratory disease should be considered. If applicable, seek follow up care with the primary health care provider.

All negative results should be treated as presumptive and confirmation a molecular assay may be necessary if there is a high likelihood of SARS-CoV-2 infection, such as in an individual with a close contact with COVID-19 or with suspected exposure to COVID-19 or in communities with high prevalence of infection. Negative results do not rule out SARS-CoV-2 infection and should not be used as the sole basis for treatment or patient management decisions, including infection control decisions.

Invalid test results

If an issue occurs a message will be displayed on the Instrument include useful information and are highlighted by an orange banne Error messages also include a A description of the Instrument status or error and an instruction. Error code that may be used for furth troubleshooting purposes. Refer to the LumiraDx Platform User Manua if an error message is displayed on the LumiraDx Instrument touch Customer Services on telephone

If the On Board Control (OBC) fails, an error message will be shown and

no test result will be returned. Follow the on screen instructions to dispose of the Test Strip and start a new test with a new Test Strip and a new drop of acted sample. If the problem persists, contact Customer Services.

if the strip has exceeded the expiry date for use, and if the strip Lot

several quality control functions integrated to ensure validity of each test run.These checks ensure that the volume of sample added is sufficient and the assay sequence of the Test Strip is as expected The checks also ensure

The LumiraDy Instrument and LumiraDy SADS CoV 2 Ad Test Strips have

that the Test Strip has not been damaged or used previously If these checks are not verified, the test run will be rejected and an error message displayed The LumiraDx Instrument ensures the quality of test results obtained through

the following features: Automated checks of the correct functioning of the Instrument at

- power on and during operation. This includes electrical component operation, heater operation,
- pattery charge state, mechanical actuators and sensors and optical system porformance
- Monitoring of Test Strip performance and controls during test runtime Ability to perform Quality Control Tests using LumiraDx Quality Control solutions to meet regulatory compliance requirements.

External Quality Controls

External liquid Quality Controls for SAPS-CoV-2 Ag are available from LumiraDx and may be used to demonstrate that the Test is functioning properly by demonstrating the expected Quality Control results and con test performance by the operator External Quality Control requirements should be established in accordance with local state, and federal regulations or accreditations requirements. It is recommended that external control testing he performed with each new operator and before using a new lot or shipment of the LumiraDx SARS-CoV-2 Ag Test. Refer to the LumiraDx SARS-CoV-2 Aa Quality Controls pack insert available at lumirady com for detailed instructions LumiraDx SARS-CoV-2 Ag Quality Controls are purchased separately.

If the LumiraDy SAPS-CoV-2 Aa Quality Controls do not perform as expected repeat the QC Test and if the problems persists, do not report patient results contact LumiraDx Customer Services on telephone number 1-888-

Cleaning and disinfection Cleaning and disinfection of the Instrument should follow and be performed

according to established site protocols and schedules. To clean the Instrument wine the external surfaces with a soft slightly damp cloth when it appears visibly dirty

It is recommended to disinfect the Instrument if contamination is suspected and at least once per day when in use with LumiraDx approved materials. Details of LumiraDx approved disinfectant materials can be found at umiraDx com. Use the material until the surface of the Instrument is visib wet Allow the surface to remain wet for 1 minute and let air dry Avoid USB ports and power inlet. Do not spray or pour solution directly onto the

This test detects both viable (live) and non-viable SARS-CoV and SARS-CoV-2 Test performance depends on the amount of virus (antigen) in the sample and may or may not correlate with viral culture results performed on the same sample. Failure to follow the instructions for use may adversely affect test

Instrument. Do not put any objects or cleaning materials into the Test Strip

- performance and/or invalidate the test result Incorrect test results may occur if a specimen is incorrectly collected
- Test results should be considered in the context of all available clinical and diagnostic information, including patient history and other test requite Positive test results do not differentiate between SARS-CoV and
- SARS-CoV-2. Negative test results are not intended to rule in other non-SARS viral or bacterial infections
- All COVID-19 antigen test negative results are presumptive and confirmation with a molecular assay may be necessary. If the patient continues to have symptoms of COVID-19, and both
- have COVID-19, however additional follow-up may be needed. If the test is positive then proteins from the virus that causes COVID-19 have been found in the sample and the individual likely nas COVID-19

the patient's first and second tests are negative, the patient may not

- The performance of this test was established based on the evaluation of a limited number of clinical specimens collected between June 2020 and March 2021. The clinical performance ha not been established in all circulating variants but is anticipated to be reflective of the prevalent variants in circulation at the time and ocation of the clinical evaluation. Performance at the time of testing may vary depending on the variants circulating, including newly emerging strains of SARS-CoV-2 and their prevalence, which change
- If the differentiation of specific SARS viruses and strains is needed additional testing, in consultation with state or local public health departments, is required. Clinical performance was established on frozen samples and
- performance may be different with fresh clinical samples. Users should test samples as quickly as possible after sample
- Extracted anterior nasal samples or nasopharynaeal samples may be frozen at -80°C and used up to 5 days after freezing. Swab samples and Extraction buffer must be at room temperature

ollected inappropriately, therefore a negative test result does not

Positive test results do not rule out co-infection with other pathogens A false negative result may occur if the level of viral antigen in a sample is below the detection limit of the test or if the sample was

rule out the possibility of SARS-CoV-2 infection.

- The amount of antigen in a sample may decrease as the duration of illness increases. Samples collected after 12 days are more likely to be negative compared to RT-PCR The contents of this kit are for qualitative detection of SARS-CoV-2
 - antigens from nasal swab and nasopharyngeal samples only. For information on swabs that have been validated by LumiraDx for
- use with the LumiraDx SARS-CoV-2 Ag Test please visit lumiradx con There is a higher change of false negative results with antigen tests than with laboratory-based molecular tests . This means that there a higher chance this test will give a negative result in an individual

with COVID-19 as compared to a molecular test especially in samples with low viral load Conditions of Authorization for the Laboratory

The LumiraDy SAPS-CoV-2 Ag Test Letter of Authorization, along with the authorized Eact Sheet for Healthcare Providers the authorized Eact Sheet for

Patients, and authorized labeling are available on the FDA website: https://www.fda.gov/medical-devices/coronavirus-disease-2019-covid

19-emergency-use-authorizations-medical-devices/in-vitro-diagnostics-However to assist clinical laboratories using the LumiraDx SARS-CoV-2 Ac

Test ("your product" in the conditions below), the relevant Conditions of Authorization are listed helow: Authorized laboratories? using your product must include with

test result reports, all authorized Fact Sheets. Under exigent

- circumstances, other appropriate methods for disseminatina this labeling may be used, which may include mass media. Authorized laboratories using your product must use your product as
- outlined in the authorized labeling. Deviations from the authorized procedures including the authorized instruments authorized clinical specimen types, authorized control materials, authorized ancillary reagents and authorized materials required to use your product are not permitted Authorized laboratories that receive your product must notify the
- relevant public health authorities of their intent to run your product prior to initiating testing. Authorized laboratories using your product must have a process in place for reporting test results to healthcare providers and relevant public health authorities, as appropriate.
- Authorized laboratories must collect information on the performance of your product and report to DMD/OHT7-OIR/OPEQ/CDRH (via email: CDRH-EUA-Reporting@fda.hhs.gov) and you (via email: customerservices. US@lumiradx.com) any suspected accurrence of false positive or false pegative results and significant deviations from the established performance characteristics of your product of which they become aware.
- All operators using your product must be appropriately trained in performing and interpreting the results of your product use appropriate personal protective equipment when handling this ki and use your product in accordance with the authorized labeling.
- You, authorized distributors, and authorized laboratories using your product must ensure that any records associated with this EUA are maintained until otherwise notified by FDA. Such records will be made available to FDA for inspection upon request

The letter of authorization refers to "Laboratories certified under the Clinical Laboratory Improvement Amendments of 1988 (CLIA), 42 U.S.C 263a, that et the requirements to perform moderate, high or waived complexity tests This product is authorized for use at the Point of Care (POC) i.e. in patient care settings operating under a CLIA Certificate of Waiver Certificate of Compliance, or Certificate of Accreditation." as "authorized

Clinical performance - Anterior Nasal Swab

The performance of the LumiraDx SARS-CoV-2 Ag Test was established with 257 direct nasal swahs prospectively collected from individual subjects between June 2020 and July 2020 during the 2020 COVID-19 pandemic. with symptoms of COVID-19 (159) or key workers (98) at increased risk of infection. No positive results were observed from patients without symptoms or beyond 12 days of symptom onset. Dual nasal swabs were nultaneously collected and then randomly allocated to testing with the umiraDy test or an ELIA PT-PCP assay Samples were collected from 6 sites across the United States (5) and United Kinadom (1), including four sites in which minimally trained operators collected and tested fresh samples.

Swabs were collected and extracted into the LumiraDx extraction buffer without transport media. Samples were tested fresh or frozen within 1h of collection and stored until tested. Samples were thawed and sequentially ted according to the Product Insert, with operators blinded to the RT-PCI result. The performance of the LumiraDx SARS-CoV-2 Aa Test was compared to the results from anterior nasal swabs collected into 3ml universal transport medium (UTM) and tested with an EUA RT-PCR method.

The Instrument reads the 2D bar code on each Test Strip and can identify

Patient demographics (age time elapsed since onset of symptoms) are available for the 257 samples used in the study. The table below shows the positive results broken down by age of the patient.

Age	LumiraDx SARS-CoV-2 Ag (n = 81)						
	Total #	Positive	Prevalence				
≤ 5 years	13	0	N/A				
6 to 21 years	29	6	20.7%				
22 to 59 years	200	70	35.0%				
≥ 60 years	15	5	33.3%				

Days since symptom onset	Cumulative RT-PCR Positive(+)	Cumulative LumiraDx Positive(+)	PPA	95% Confidence interval		
0	6	6	100.0%	61.0%	100.0%	
1	12	12	100.0%	75.8%	100.0%	
2	28	28	100.0%	87.9%	100.0%	
3	37	37	100.0%	90.6%	100.0%	
4	55	54	98.2%	90.4%	99.7%	
5	61	60	98.4%	91.3%	99.7%	
6	67	66	98.5%	92.0%	99.7%	
7	73	73 72		92.6%	99.8%	
8	75	74	98.7%	92.8%	99.8%	
9	75	74	98.7%	92.8%	99.8%	
10	77	76	98.7%	93.0%	99.8%	
11	80	79	98.8%	93.3%	99.8%	
12	83	81	97.6%	91.6%	99.3%	

Final data analysis is presented below

Reference	e RT-PCR	Assay					95% Wilson Score CI		
							LCI	UCI	
		POS	NEG	Total	PPA	97.6%	91.6%	99.3%	
LumiraDx	POS	81	6	87	NPA	96.6%	92.7%	98.4%	
SARS-CoV-2 Ag Test	NEG	2	168	170	Prevalence	32.3%	26.9%	38.2%	
	TOTAL	83	174	257					

PPA - Positive Percent Agreement (Sensitivity)

NPA - Negative Percent Agreement (Specificity)

LCI - Lower Confidence Interval

UCI - Upper Confidence Interval

The performance of the SARS-CoV-2 Ag Test was established with 255 nasopharyngeal swabs prospectively collected from individual subjects between August 2020 and September 2020 during the 2020 COVID pandemic. Subjects were presenting with symptoms of COVID-19 being screened for infection. Samples were collected from 6 sites across the United States. Swabs were collected and extracted into the LumiraDx Extraction Buffer. Samples were tested fresh within 1h of collection and tested according to the Product Insert. The performance of the LumiraDx SARS-COV-2 Ag Test was compared to the results from nasopharyngeal samples collected into 3ml universal transport medium (UTM) and tested with an EUA authorized PCR method.

Patient demographics (age, time elapsed since onset of symptoms) are available for the 255 samples used in the study. The table below shows the positive results broken down by age of the patient:

Age	Lu	LumiraDx SARS-CoV-2 Ag (n = 39)						
	Total #	Positive	Prevalence					
≤ 5 years	22	0	0.0%		2			
6 to 21 years	59	9	15.3%					
22 to 59 years	150	28	18.7%		4			
≥ 60 years	24	2	8.3%		6			

									Limit of Detection - LOD (analytical
Days since symptom onset	Cumulative PCR Positive (+)	LumiraDx Positive (+)	PPA	LCI	UCI	NPA	LCI	UCI	Limit of Detection (LoD) studies det The LoD for the LumiraDx SARS-CoV
0	2	2	100.0%	34.2%	100.0%	100.0%	75.8%	100.0%	52287 is a preparation of SARS-Rela The material was supplied frozen a
1	6	6	100.0%	61.0%	100.0%	100.0%	93.4%	100.0%	Limit of Detection (LoD) screening
2	9	9	100.0%	70.1%	100.0%	100.0%	96.2%	100.0%	An initial LoD screening study was p
3	17	17	100.0%	81.6%	100.0%	98.6%	94.9%	99.6%	human nasal matrix starting at a te These dilutions were tested in triplic
4	22	22	100.0%	85.1%	100.0%	98.8%	95.7%	99.7%	were positive was chosen for LoD R
5	23	23	100.0%	85.7%	100.0%	98.4%	95.3%	99.4%	1.6 TCID ₅₀ /swab.
6	26	26	100.0%	87.1%	100.0%	98.5%	95.6%	99.5%	SARS-CoV-2
7	34	34	100.0%	89.8%	100.0%	98.5%	95.7%	99.5%	
8	36	36	100.0%	90.4%	100.0%	98.6%	95.8%	99.5%	
9	36	36	100.0%	90.4%	100.0%	98.6%	95.9%	99.5%	
10	39	38	97.4%	86.8%	99.5%	98.1%	95.2%	99.3%	
11	40	39	97.5%	87.1%	99.6%	97.7%	94.6%	99.0%	
12	40	39	97.5%	87.1%	99.6%	97.7%	94.7%	99.0%	

Reference	e RT-PCR	Assay					95% Wilson Score CI		
							LCI	UCI	
		POS	NEG	Total	PPA	97.5%	87.1%	99.6%	
LumiraDx	POS	39	5	44	NPA	97.7%	94.7%	99.0%	
SARS-CoV-2 Ag Test	NEG	1	210	211	Prevalence	15.7%	11.7%	20.7%	
	TOTAL	40	215	255					

NIH study on the A prospective clinical study was conducted between January 2021 and May 2022 as a component of the Panid Acceleration of Diagnostics (PADy) initiative

from the National Institutes of Health (NIH). A total of 7,361 individuals were enrolled via a decentralized clinical study design, with a broad geographical representation of the United States. Per inclusion criteria, all inclividuals were asymptomatic upon enrollment in the study and at least 14 days prior to it and did not have a SARS-CoV-2 infection in the three months prior to enrollment. Participants were assigned to one of three ELIA authorized SARS-CoV-2 OTC rapid antigen tests to conduct serial testing (every 48 hours) for 15 days If an antigen test was positive the serial-antigen testing result is considered positive At each rapid antigen testing time point, study subjects also collected a nasal swab for comparator testing using a home collection kit (using a 15-minute normalization window between swabs). SARS-CoV-2 infection status was determined by a composite comparator method on the day of the first antigen test, using at least two highly sensitive EUA RT-PCRs. If results of the first two molecular test were discordant a third highly sensitive EUA RT-PCR test was performed,

Study participants reported symptom status throughout the study using the MyDataHelps app. Two-day serial antiqen testing is defined as performing two antigen tests 36 - 48 hours apart. Three-day serial antigen testing is defined as performing three antigen tests over five days with at least 48 hours between

Out of the 7.361 participants enrolled in the study 5.609 were eliable for analysis. Among eliable participants, 154 tested positive for SARS-CoV-2 infection based on RTPCR, of which 97 (62%) were asymptomatic on the first day of infection, whereas 57 (39%) reported symptoms on the first day of infection. Pre-symptomatic subjects were included in the positive percent agreement (PPA) of asymptomatic individuals, if they were asymptomatic on the first day of antigen testing, regardless of whether they developed symptoms at any time after the first day of testing. Performance of the antigen test with serial testing in individuals is described in the table below.

Note: This study was not performed using LumiraDx assay.

and the final test result was based upon the majority rule

Data establishing PPA of COVID-19 antigen serial testing compared to the molecular comparator single day testing throughout the course of infection with serial testing. Data is from all antigen tests in the study combined (LumiraDx not included in this study), however, the data is being applied to all antigen test

ys after first PCR sitive Test Result	Asympto	omatic on first day of	testing	Symptomatic on first day of testing				
		Ag Positiv	e / PCR Positive (An	tigen Test Performand	ce % PPA)			
	1 Test	2 Tests	3 Tests	1 Test	2 Tests	3 Tests		
0	9/97	35/89	44/78	34/57	47/51	44/47		
	(9.3%)	(39.3%)	(56.4%)	(59.6%)	(92.2%)	(93.6%)		
2	17/34	23/34	25/32	58/62	59/60	43/43		
	(50.0%)	(67.6%)	(78.1%)	(93.5%)	(98.3%)	(100%)		
4	16/21	15/20	13/15	55/58	53/54	39/40		
	(76.2%)	(75.0%)	(86.7%)	(94.8%)	(98.1%)	(97.5%)		
6	20/28	21/27	16/18	27/34	26/33	22/27		
	(71.4%)	(77.8%)	(88.9%)	(79.4%)	(78.8%)	(81.5%)		
8	13/23	13/22	4/11	12/17	12/17	7/11		
	(56.5%)	(59.1%)	(36.4%)	(70.6%)	(70.6%)	(63.6%)		
10	5/9 (55.6%)	5/8 (62.5%)	-	4/9 (44.4%)	3/7 (42.9%)	-		

2 Tests = two (2) tests performed an average of 48 hours apart. The first test performed on the indicated day and the second test performed 48 hours later. 3 Tests = three (3) tests performance an average of 48 hours apart. The first test performed on the indicated day, the second test performed 48 hours later, and a final test performed 48 hours after the second test.

1 Test = one (1) test performed on the noted days after first PCR positive test result. Day 0 is the first day of documented infection with SARS-CoV-2.

ositive and negative re	sults broken down by days	since symptom onset						Analytical performance					
Days since symptom onset	Cumulative PCR Positive (+)	LumiraDx Positive (+)	PPA	LCI	UCI	NPA	LCI	UCI	Limit of Detection - LOD (analytical sensitivity) Limit of Detection (LoD) studies determined the lowest detectable concentration of SARS-CoV-2 at which 100% of all (true positive) replicates test positive. The LoD for the LumiraDx SARS-CoV-2 Ag Test was established using limiting dilutions of gamma-irradiated SARS-CoV-2 (BEIResources NR-52287). The NR-52287 is a preparation of SARS-Related Coronavirus 2 (SARS-CoV-2), isolate USA WA1/2020, that has been inactivated by gamma-irradiation at 5 x 10° RADs. The material was supplied frozen at a concentration of 2.8 x 10° TCID.,/mL.				
0	2	2	100.0%	34.2%	100.0%	100.0%	75.8%	100.0%					
1	6	6	100.0%	61.0%	100.0%	100.0%	93.4%	100.0%	Limit of Detection (LoD) screening				
2	9	9	100.0%	70.1%	100.0%	100.0%	96.2%	100.0%	An initial LoD screening study was performed using a 5-fold serial dilutions (six dilutions in total) of the gamma-irradiated virus made in pooled negative				
3	17	17	100.0%	81.6%	100.0%	98.6%	94.9%	99.6%	human nasal matrix starting at a test concentration of 2 x 10°TCID _{ss} /mL (as shown in table below) and processed for each study as described above. These dilutions were tested in triplicate. The volume of solution spiked onto each swab was 50µL. The lowest concentration at which all (3 out of 3 replicates				
4	22	22	100.0%	85.1%	100.0%	98.8%	95.7%	99.7%	were positive was chosen for LoD Range finding. This was 32 TCID _{sn} /mL. Based upon the testing procedure for this study the LoD of 32 TCID _{sn} /mL equates to				
5	23	23	100.0%	85.7%	100.0%	98.4%	95.3%	99.4%	1.6TCID ₅₀ /swab.				
6	26	26	100.0%	87.1%	100.0%	98.5%	95.6%	99.5%	SARS-CoV-2 tested (TCID _{so} /mL)	Test result			
7	34	34	100.0%	89.8%	100.0%	98.5%	95.7%	99.5%	20000	3/3 positive			
8	36	36	100.0%	90.4%	100.0%	98.6%	95.8%	99.5%	4000	3/3 positive			
9	36	36	100.0%	90.4%	100.0%	98.6%	95.9%	99.5%	800	3/3 positive			
10	39	38	97.4%	86.8%	99.5%	98.1%	95.2%	99.3%	160	3/3 positive			
11	40	39	97.5%	87.1%	99.6%	97.7%	94.6%	99.0%	32	3/3 positive			
	40		07.50	07.10	00.404	07.70	0.4.70/	00.00/	6.4	0/2 positivo			

Using the 32 TCID.../mL concentration, the LoD was further refined using a 2-fold dilution series (four dilutions in total) of the gamma-irradiated SARS-CoV-2 using in the 22 - 10-20/THZ concentration, in the 20 was taking in the 12 - 10-20 attained some state of the 10-20 attained and the 10-20 positive was treated as the tentative LoD for the LumiraDx SARS-CoV-2 Ag Test. This was 32 TCID_{so}/mL.

							LCI	UCI			
		POS	NEG	Total	PPA	97.5%	87.1%	99.6%			
aDx 2 Ag Test	POS	39	5	44	NPA	97.7%	94.7%	99.0%			
	NEG	1	210	211	Prevalence	15.7%	11.7%	20.7%			
	TOTAL	40	215	255							
hree Lateral I	ree Lateral Flow OTC rapid Antigen tests (Not including LumiraDx)										

Limit of Detection (LoD) confirmation

SARS-CoV-2 tested (TCID_{co}/mL)

The LoD of the LumiraDx SARS-CoV-2 Ag Test was then confirmed by testing 20 replicates with concentrations at the tentative Limit of Detection. The final LoD of the LumiraDx SARS-CoV-2 Ag Test was determined to be the lowest concentration resulting in positive detection of twenty (20) out of twenty (20) replicates. Based on this testing the LoD for nasal swab samples was confirmed as: 32 TCID_{en}/mL. Based upon the testing procedure for this study the LoD of 32 TCID_{en}/mL eauates to 1.6 TCID // swab.

Starting Material Concentration	Estimated LOD	No. Positive/Total	% Positive	
2.8 x 10 ⁵ TCID ₅₀ /mL	32 TCID ₅₀ /mL	20/20	100	

Test result

3/3 positive 0/3 positive

1/3 positive

0/3 positive

performance of this test device in the detection of the Omicron variant of SARS-CoV-2 was evaluated in a dilution series of clinical specimens which

were positive for the Omicron variant This testing was conducted by the National Institutes of Health (NIH) as a component of the Rapid Acceleration of oliganostics (RADx®) initiative. Specimen pools were prepared by the RADx® team using clinical pooled samples from currently circulating Omicron strains tested by RADX® to assess performance with the Omicron variant. Results from this dilution series cannot be compared to other specimen pools and do at indicate that a test will have different clinical performance compared to other FLIA authorized tests. Compared to an FLIA-authorized PTPCP method this SARS-CoV-2 Aa test detected 100% of live virus Omicron samples at a Ct-value of 23.6 (n=5). Less than 50% of Omicron dilutions at Ct-values of 24.0 and 24.8 were detected (n=5). Omicron dilutions at lower viral concentrations (Ct-values greater than 25.8 for live virus) were not detected by LumiraDx SARS-CoV-2 Ag Test in this study. See summary data table below.

Omicron Pool 2 - Live Dilution		Assay #1	Assay #2	LumiraDx SARS-CoV-2 Ag Test*	
	Ct-N2 Ave.	Percent Positive (n=5)	Percent Positive (n=5)	Percent Positive (n=5)	
Dilution 1	19.8	100	100	100	
Dilution 2	20.8	100	100	100	
Dilution 3	21.5	100	100	100	
Dilution 4	22.7	100	100	100	
Dilution 5	23.6	100	0	100	
Dilution 6	24.0	60	0	40	
Dilution 7	24.8	0	0	20	
Dilution 8	25.8	0	0	0	
Dilution 9	27.4	0	0	0	
Dilution 10	28.1	0	0	0	
Dilution 11	29.1	0	0	0	

esting was conducted using a multiplexed version of the test, which comprises the same SARS-CoV-2 test design and components.

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Cross-reactivity (analytical specificity) and microbial interference studies

Microorganism

Human coronavirus 220E

Cross-reactivity and inteference of the LumiraDx SARS-CoV-2 Ag Test was evaluated by testing a panel of related pathogens, high prevalence disease agents and normal or pathogenic flora including various microorganisms and viruses and populitive matrix that are regardably likely to be encountered in the clinical sample and could potentially cross-react or interfere with the LumiraDx SARS CoV-2 Aa Test. Each organism and virus were tested in the absence or ance of heat inactivated SARS-CoV-2 at 3 v LoD

Cross-Reactivity (Yes/No)

No (3/3 negative)

Interference (Yes/No)

No. (3/3 positive)

Concentration

Source

Dexc	. (.,.,.,,	ito (o/o nogamo)			
	No (19/20 positive)	uman coronavirus OC43 Zeptometrix 1 x 10 ⁵ PFU/mL No (3/3 negative) No (Human coronavirus OC43	
	No (3/3 positive)	No (3/3 negative)	9.87 x 10 ³ PFU/mL	Zeptometrix	Human coronavirus NL63
High dose hook effect	No (3/3 positive)	No (2/2 negative)	7930 PFU/mL	Zeptometrix	MERS coronavirus
High Dose Hook Effect study To determine if the Lumira	No (3/3 positive)	No (3/3 negative)	1 x 10 ⁵ PFU/mL	Zeptometrix	Adenovirus (e.g. C1 Ad. 71)
Resources NR-52287) were	No (3/3 positive)	No (3/3 negative)	1 x 10 ⁵ PFU/mL	Zeptometrix	Human Metapneumovirus (hMPV)
nasal matrix obtained fror processed for testing on the	No (3/3 positive)	No (3/3 negative)	1 x 10 ⁵ PFU/mL	Zeptometrix	Parainfluenza virus Type 1
No impact on test perform	No (3/3 positive)	No (3/3 negative)	1 x 10 ⁵ PFU/mL	Zeptometrix	Parainfluenza virus Type 2
CoV-2 Ag Test.	No (3/3 positive)	No (3/3 negative)	1 x 10 ⁵ PFU/mL	Zeptometrix	Parainfluenza virus Type 3
	No (3/3 positive)	No (3/3 negative)	1 x 10 ⁵ PFU/mL	Zeptometrix	Parainfluenza virus Type 4a
Test dilutio	No (3/3 positive)	No (3/3 negative	8.82 x 10 ⁴ PFU/mL	Zeptometrix	Influenza A H3N2 (Wisconsin/67/05)
1	No (3/3 positive)	No (3/3 negative)	1 x 10 ⁵ PFU/mL	Zeptometrix	Influenza A H1N1
2	No (19/20 positive)	No (3/3 negative)	2.92 x 10 ⁴ PFU/mL	Zeptometrix	Influenza B (Malaysia/2506/04)
3	No (3/3 positive)	No (3/3 negative)	1 x 10 ⁵ PFU/mL	Zeptometrix	Enterovirus
4	No (3/3 positive)	No (3/3 negative)	1 x 10 ⁵ PFU/mL	Zeptometrix	Respiratory syncytial virus
5	No (3/3 positive)	No (3/3 negative)	4.17 x 10 ⁵ PFU/mL	Zeptometrix	Rhinovirus
	No (3/3 positive)	No (3/3 negative)	1 x 106 CFU/mL	Zeptometrix	Haemophilus influenzae
Point of care use	No (3/3 positive)	No (3/3 negative)	1 x 106 CFU/mL	Zeptometrix	Streptococcus pneumoniae
The LumiraDx SARS-CoV-2	No (3/3 positive)	No (3/3 negative)	1 x 106 CFU/mL	Zeptometrix	Streptococcus pyogenes
References	No (3/3 positive)	No (3/3 negative)	1 x 106 CFU/mL	Zeptometrix	Candida albicans
 World Health Orga 	No (3/3 positive)	No (3/3 negative)	14% v/v	LumiraDx	Pooled human nasal wash
Centers for Disease	No (3/3 positive)	No (3/3 negative)	1 x 106 CFU/mL	Zeptometrix	Bordetella pertussis
Symbols glossary	No (3/3 positive)	No (3/3 negative)	1 x 106 CFU/mL	ATCC	Mycoplasma pneumoniae
1c	No (3/3 positive)	No (3/3 negative)	1 x 106 CFU/mL	ATCC	Chlamydia pneumoniae
Temperature	No (3/3 positive)	No (3/3 negative)	1 x 106 CFU/mL	Zeptometrix	Legionella pneumophila
	No (3/3 positive)	No (3/3 negative)	1 x 106 CFU/mL	Zeptometrix	Mycobacterium tuberculosis
Manufacture	No (3/3 positive)	No (3/3 negative)	1 x 106 CFU/mL	Zeptometrix	Pneumocystis jirovecii
[12.50]	No (3/3 positive)	No (3/3 negative)	1 x 106 CFU/mL	Zeptometrix	Pseudomonas Aeruginosa
IVD In Vitro Diagn	No (3/3 positive)	No (3/3 negative)	1 x 106 CFU/mL	Zeptometrix	Staphylococcus Epidermidis
	No (3/3 positive)	No (3/3 negative)	1 x 106 CFU/mL	Zeptometrix	Streptococcus Salivarius
REF Catalogue N	No (3/3 positive)	No (3/3 negative)	1 x 106 CFU/mL	ATCC	Staphylococcus aureus

Alignment Search Tool (BLAST) managed by the National Center for Biotechnology Information (NCBI) was used to assess the degree of protein sequence For Human Coronavirus HKU1, homoloay exists between the SARS-CoV-2 nucleocapsid protein and Human Coronavirus HKU1. BLAST results showed 30

homologous across 76% of the sequences, this is relatively low but cross-reactivity cannot be fully ruled out. For SARS-Coronavirus, high homology exists between the SARS-CoV-2 nucleocapsid protein and SARS-Coronavirus, BLAST results showed 68 sequence IDs, mostly nucleocapsid protein, showing homology. Sequence ID AAR87518.1, had the highest alignment score isolated from a human patient and was found to be 90.76% homologous across 100% of the sequence. This is high and cross-reactivity is likely.

sequence IDs, all nucleocapsid protein, showing homology. Sequence ID AGW27840.1 had the highest alignment score and was found to be 39.1%

For MERS-Coronavirus, high homology exists between the SARS-CoV-2 nucleocapsid protein and MERS-Coronavirus. BLAST results showed at least 114 sequence IDs, mostly nucleocapsid protein, showing homology. Sequence IDs AHY61344.1 and AWH65950.1, had the highest alignment scores isolated from a human patient and were found to be 49.4% and 50.3% homologous across 88% of the sequence. Whilst this potentially represents moderate cross-reactivity testing of the MERS virus at 7930 PFU/mL showed no reactivity (see table above).

Endogenous interference studies

A study was performed to demonstrate that potentially interfering substances that may be found in the upper respiratory tract in symptomatic subjects (including over the counter medications) do not cross-react or interfere with the detection of SARS-CoV-2 in the LumiraDx SARS-CoV-2 Ag Test. Each substance was tested in triplicate in the absence or presence of SARS-CoV-2 at 3 x LoD. Substances for testing were selected based on the respiratory samples guidance in http://www.accessdata.fda.gov/cdrh_docs/reviews/K112177.pdf. The final concentration of the substances tested are documented in the Table below.

Interfering substance	Concentration	Interference (Yes/No)
Benzocaine	150 mg/dL	No (3/3 Negative, 3/3 Positive)
Blood (human)	5%	No (3/3 Negative, 3/3 Positive)
Mucin	5 mg/mL	No (3/3 Negative, 3/3 Positive)
Naso GEL (NeilMed)	5% v/v	No (3/3 Negative, 3/3 Positive)
CVS Nasal Drops (phenylephrine)	15% v/v	No (3/3 Negative, 3/3 Positive)
Afrin (Oxymetazoline)	15% v/v	No (3/3 Negative, 3/3 Positive)
CVS Nasal Spray (Cromolyn)	15% v/v	No (3/3 Negative, 3/3 Positive)
Zicam Cold Remedy	5% v/v	No (3/3 Negative, 3/3 Positive)
Homeopathic (Alkalol)	10 % v/v	No (3/3 Negative, 3/3 Positive)
Sore Throat Phenol Spray	15% v/v	No (3/3 Negative, 3/3 Positive)
Tobramycin	3.3 mg/dL	No (3/3 Negative, 3/3 Positive)
Mupirocin	0.15 mg/dL	No (3/3 Negative, 3/3 Positive)
Fluticasone	0.000126 mg/dL	No (5/5 Negative, 4/4 Positive)
Tamiflu (Oseltamivir phosphate)	500 mg/dL	No (3/3 Negative, 3/3 Positive)
Budesonide	0.00063 mg/dL	No (3/3 Negative, 3/3 Positive)
Biotin	0.35 mg/dL	No (3/3 Negative, 3/3 Positive)

Concentration Interfering substance Interference (Ves/No) Methanol 150 ma/dL No (19/20 Negative, 3/3 Positive) Acetylsalicylic Acid 3 ma/dL No (3/3 Negative, 3/3 Positive) 0.0774 ma/dL Diphenhydramine No (3/3 Negative, 3/3 Positive) 0.00156 mg/dL No (19/20 Negative, 3/3 Positive) Dextromethornhan No (3/3 Negative, 3/3 Positive) Devamethasone 1.2 mg/dL No (3/3 Negative, 3/3 Positive) Muciney

ah Dose Hook Effect studies determine the level at which false negative results can be seen when very high levels of target are present in a tested sample

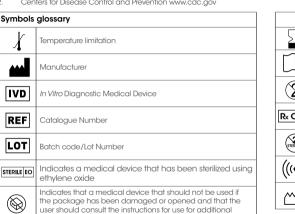
ight buse how. Each studies deleth line hie level of which has hegalities lessels with very high levels of halper are plesen in the lessed sample. to determine if the LumiraDx SARS-CoV-2 Ag Test suffers from any high dose hook effect, increasing concentrations of gamma-irradiated SARS CoV-2 virus (BEI esources NR-52287) were tested up to a concentration of 1.4 x 10° TCID₂₀/mL. In this study, the starting material was spiked into a volume of pooled human asal matrix obtained from healthy donors and confirmed negative for SARS-CoV-2. At each dilution, 50 µL samples were added to swabs and the swabs ocessed for testing on the LumiraDx SARS-CoV-2 Ag Test as per the Product Insert using the procedure appropriate for patient nasal swab samples. o impact on test performance or high dose hook effect was observed up to 1,4 x 10°TCID.../mL of gamma-irradiated SARS-CoV-2 with the LumiraDx SARS-

Test dilution	Concentration (TCID50/mL)	Mean signal (ADC Units)
1	0	495
2	62.5	26100.6
3	250	63013.8
4	1000	83451.8
5	1.4 x 10 ⁵	86220

ne LumiraDx SARS-CoV-2 Ag Test was used by 8 untrained users in 4 sites across the United States. Untrained users tested 132 patients and ran 148 tests.

World Health Organisation www.who.int

Centers for Disease Control and Prevention www.cdc.gov



Consult Instructions for Use 2 Do Not Re-use Rx Only Prescription Use Only STEENER Do not re-sterilize ((•))) Indicates the presence of model in Identification (RFID) reader/tag. Indicates the presence of the Radio Frequency Date of manufacture

LumiraDx customer services:

For product inquiries and technical support please contact LumiraDx Customer Services by email: customerservices.US@lumiradx.com, telephone 1-888-586-4721 or Lumiradx.com

If there is a problem with the LumiraDx SARS-CoV-2 Ag Test Strips you may be asked to return them. Before returning tests please obtain a return authorization number from LumiraDx Customer Services. This return authorization number must be on the shipping carton for return. For ordinary returns following purchase, please contact LumiraDx Customer Services for terms and conditions; customerservices.US@lumiradx.com

Limited warranty

LumiraDx SARS-CoV-2 Ag Test Strips – As per shelf life.

Unused strips and nasal collection swabs must be stored according to the required storage conditions as printed in this product insert and they can be used only up to the expiry date printed on the Test Strip pouch, Test Strip box and swab packaging. For the applicable warranty period, LumiraDx warrants that each product shall be (1) of good quality and free of material defects, (ii) function in accordance with the material specifications referenced in the product insert, and (iii) approved by the proper governmental agencies required for the sale of products for their intended use (the "limited warranty"). If the product fails to meet the requirements of the limited warranty, then as customer's sole remedy, LumiraDx shall either repair or replace, at LumiraDx's discretion, the Test Strips. Except for the limited warranty stated in this section, LumiraDx disclaims any and all warranties, express or implied, including but not limited to, any warranty of merchantability, fitness for a particular purpose and non-infringement regarding the product. LumiraDx's maximum liability with any customer claim shall not exceed the net product price paid by the customer. Neither party shall be liable to the other party for special, incidental or consequential damages, including, without limitation, loss of business, profits, data or revenue, even if a party receives notice in advance that these kinds of damages migh result. The Limited Warranty above shall not apply if the customer has subjected the LumiraDx SARS-CoV-2 Ag Test to physical abuse, misuse, abnormal use, use inconsistent with the LumiraDx Platform User Manual or Product Insert, fraud, tampering, unusual physical stress, negligence or accidents. Any warranty claim by Customer pursuant to the Limited Warranty shall be made in writing within the applicable Limited Warranty period.

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Manufacturer information

Test Strips

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